Evolution and spread of multi-adapted pathogens in a spatially heterogeneous environment

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Abstract

Pathogen adaptation to multiple selective pressures challenges our ability to control their spread. 2 Here we analyse the evolutionary dynamics of pathogens spreading in a heterogeneous host popu-3 lation where selection varies periodically in space. We study both the transient dynamics taking place at the front of the epidemic and the long-term evolution far behind the front. We identify five types of epidemic profiles arising for different levels of spatial heterogeneity and different costs of adaptation. In particular, we identify the conditions where a generalist pathogen carrying multiple adaptations can outrace a coalition of specialist pathogens. We also show that finite host populations promote the spread of generalist pathogens because demographic stochasticity enhances 9 the extinction of locally maladapted pathogens. But higher mutation rates between genotypes can 10 rescue the coalition of specialists and speed up the spread of epidemics for intermediate levels of 11 spatial heterogenity. Our work provides a comprehensive analysis of the interplay between migra-12 tion, local selection, mutation and genetic drift on the spread and on the evolution of pathogens 13 in heterogeneous environments. This work extends our fundamental understanding of the outcome 14 of the competition between two specialists and a generalist strategy (single-versus multi-adapted 15 pathogens). These results have practical implications for the design of more durable control strate-16 17 gies against multi-adapted pathogens in agriculture and in public health.

Impact summary: Pathogen adaptation is constantly eroding the efficacy of prophylactic and therapeutic measures against the spread of infectious diseases. A promising way to limit the spread of multi-adapted pathogens is to distribute different control measures across space (e.g., different vaccines, different resistant varieties of crop in agriculture). Yet, the influence of the spatial deployment of these interventions on the genetic composition of spreading epidemics remains unclear. Is it possible to identify optimal deployment strategies that reduce the spread and the speed of adaptation of resistant pathogens? We analyse the evolution of pathogen adaptations throughout an epidemic spreading in a heterogeneous host population where selection varies periodically in space. We show how lower spatial heterogeneity can speed up the epidemic spread and disfavour multi-adapted pathogens. But this effect can be altered qualitatively by the demographic stochasticity taking place at the edge of the front and by higher rates of mutation between different pathogen genotypes. We predict the composition of the pathogen population both *far behind* and *at* the front of the epidemic. This analysis allows us to elucidate the consequences of the effects of spatial heterogeneity on the coexistence between specialist (single-adaptated) and generalist (multi-adapted) pathogen strategies.

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19 1 Introduction

Pathogen epidemics can have devastating consequences for animal and plant species and it is particu-20 larly important to understand which factors govern the speed of epidemics to predict and potentially 21 prevent their spread. Determining the speed of biological invasions has attracted a lot of attention 22 from theoretical biologists (Fisher 1937; Kolmogorov, Petrovsky, and Piskunov 1937; Skellam 1951). 23 Under the simplifying assumption that the invasion takes place in a homogeneous environment (e.g. 24 an epidemic spreading in a fully susceptible host population), diffusion models can be used to pre-25 dict the asymptotic speed of the epidemic (Fisher 1937; Kot, Lewis, and Driessche 1996; Shigesada 26 and Kawasaki 1997). In this case the population is expected to spread as a travelling wave with a 27 constant speed equal to $2\sqrt{\sigma r}$, where r is the growth rate of the population at low density and σ is 28 the diffusion coefficient that measures how quickly the organisms disperse. Spatial heterogeneity in 29 the environment, however, may dramatically affect the spread of the invading organism (Shigesada 30 and Kawasaki 1997). If the spatial variation is periodic, the natural extension of the travelling front 31 is the so-called *pulsating front* characterized by its average speed (Berestycki, Hamel, and Roques 32 2005a,b; Shigesada and Kawasaki 1997). Earlier studies have mostly focused on the spatial dynamics 33 of invasions under the assumption that evolutionary dynamics could be neglected. Yet, evolution can 34 be very rapid during invasions and this evolution can affect the speed of the spread in homogeneous 35 environments (Griette, Raoul, and Gandon 2015; Osnas, Hurtado, and Dobson 2015; Perkins et al. 36 2013; Wei and Krone 2005). 37

Here we study how the pathogen evolution can affect the spread of an epidemic taking place in a 38 spatially heterogeneous host population. Host variation is assumed to affect resistance to infection and 39 pathogen transmission. Many different situations could generate this type of spatial heterogeneity. For 40 instance, in agriculture the use of different resistant varieties in crops could be a way to manipulate the 41 spatial distribution of host resistance to a specific pathogen (Gilligan 2008; Mikaberidze, McDonald, 42 and Bonhoeffer 2015; Mundt 2002). In animal species, the use of different vaccines at different loca-43 tions could also generate a spatial mosaic of immunity (McLeod, Wahl, and Mideo 2021). Crucially, 44 we allow the pathogen to adapt to this diversity of host resistance and we consider different types of 45 adaptations. First, the pathogen may evolve a specialist strategy allowing the optimal exploitation of 46 a single resistant host. Second, the pathogen may evolve a generalist strategy allowing the pathogen 47 to exploit distinct resistant hosts. But this ability to infect multiple host may carry intrinsic fitness 48 costs (e.g. a lower transmission rate). The analysis of the competition between specialist and gen-49 eralist strategies is a classical evolutionary question which has been explored by theoretical studies 50 under different biological scenarios (Levins 1968; Parvinen and Egas 2004; Wilson and Yoshimura 51

1994). These studies have shown that the long-term evolutionary outcome and the potential coexis-52 tence between multiple strategies depend on the balance between the amount of spatial heterogeneity 53 and the homogenizing effect of migration. Yet, it is unclear if the same principle holds away from 54 the equilibrium, at the front of a population that is spreading in a heterogeneous environment. In 55 particular, it is unclear if one expects the generalist strategy to be more frequent at the edge or far 56 behind the front, and how this evolution can affect the speed of the spread. Besides, a better under-57 standing of the consequences of the heterogeneity of host resistance on pathogen dynamics could have 58 practical implications for disease control. For instance, we could optimize the composition of the host 59 population to reduce epidemic spread and limit the evolution of multi-adapted pathogens which are 60 expected to erode dramatically the efficacy of control efforts. 61

In the following, we take advantage of the theoretical framework of pulsating fronts to examine 62 the spatial dynamics of different pathogens spreading in a one-dimensional environment. First, we 63 study the effect of the spatial heterogeneity on the speed of a monomorphic pathogen population. In a 64 second step, we allow mutations between different pathogen genotypes and we analyse the evolution of 65 a coalition of different pathogen genotypes. We contrast the composition of the pathogen population 66 at the edge and behind the front and we identify five different types of epidemic profiles. Finally, we 67 examine the effect of demographic stochasticity on the speed of spreading epidemics when the host 68 population is assumed to be of finite size. 69

$_{70}$ 2 Methods

We model the dynamics of a directly transmitted pathogen in a one-dimensional habitat. At time 71 and position x, the host population is divided into uninfected individuals, S(t,x), and infected t 72 individuals, I(t, x). We assume that dead hosts are immediately replaced by new susceptible hosts 73 (because host fecundity is assumed to be large and not limiting) so that the total density of hosts is 74 assumed to remain constant over space and time: K = S(t, x) + I(t, x). We focus on a scenario where 75 the environment is divided into two different habitats where the hosts are either of type A or of type 76 B. For instance, this scenario could result from the use of two different vaccines at different locations 77 or, if we consider the spread of a phytopathogen in crop, by the use of distinct host resistant varieties 78 in different fields. We consider a simple spatial pattern where host composition varies periodically and 79 we use L to denote the period of the spatial fluctuation of host composition. Because all the hosts 80 are resistant to some pathogen genotype we expect that the pathogens fully sensitive to both types 81 of host resistance will be rapidly outcompeted by single- or multi-adapted genotypes. We thus focus 82 our analysis on the dynamics of three adapted pathogen genotypes circulating in the host population: 83

(i) the density of hosts infected with the genotype only able to infect host of type A is noted $I_a(t, x)$ 84 (single-adapted genotype a to host type A), (ii) the density of hosts infected with the genotype only 85 able to infect host of type B is noted $I_b(t,x)$ (single-adapted genotype b to host type B) and (iii) 86 the density of hosts infected with the genotype able to infect both types of hosts is noted $I_m(t,x)$ 87 (m for multi-adaptation to both types of hosts). Coinfection by different genotypes is not allowed 88 and each genotype i is characterized by $\beta_i(x)$, the rate at which transmission occurs between infected 89 and susceptible hosts after a contact at position x. The rate of transmission of the multi-adapted 90 genotype β_m is independent of space because multi-adaptation implies that the rate of transmission 91 is not affected by the treatment. In contrast, the rates of transmission $\beta_a(x)$ and $\beta_b(x)$ vary in space 92 because we assume that host resistance reduces transmission (without affecting the other life history 93 traits). All the infections are assumed to end (because of clearance and/or increased mortality due 94 to pathogen virulence) at a rate α . More precisely, we assume that β_a (resp. β_b) takes values $\alpha + r$ 95 in populations of host A only (resp. B only), and value $\alpha - r$ in populations of host B only (resp. A 96 only), see **Fig. 1**. This symmetry between the two specialists simplifies the following analysis of the 97 model. Note, however, that we also examine a scenario when we introduce some asymmetry in the 98 maximal growth rates of the two specialists in the **Supplementary Information** (section 1.2.1 and 99 Fig. S3). Mutations may occur between these three genotypes and μ_{ij} stands for the rate of mutation 100 from genotype i to genotype j. 101

The transmission of the pathogen is assumed to be local (infected hosts can only infect susceptible hosts at the same spatial location) but both susceptible and infected hosts are allowed to diffuse in one dimension with a fixed rate σ . In other words, we neglect the influence the pathogen may have on the mobility of its host. Our model can thus be written as the following set of reaction-diffusion equations (for readability, we drop the time and space dependence notation on host densities):

$$\begin{cases} \frac{\partial I_a}{\partial t} = I_a \left[r_a(x) - \beta_a(x) \frac{I}{K} \right] + \sigma \frac{\partial^2 I_a}{\partial x^2} + \mu_{ba} I_b + \mu_{ma} I_m - (\mu_{ab} + \mu_{am}) I_a \\ \frac{\partial I_b}{\partial t} = I_b \left[r_b(x) - \beta_b(x) \frac{I}{K} \right] + \sigma \frac{\partial^2 I_b}{\partial x^2} + \mu_{ab} I_a + \mu_{mb} I_m - (\mu_{ba} + \mu_{bm}) I_b \\ \frac{\partial I_m}{\partial t} = I_m \left[r_m - \beta_m \frac{I}{K} \right] + \sigma \frac{\partial^2 I_m}{\partial x^2} + \mu_{am} I_a + \mu_{bm} I_b - (\mu_{ma} + \mu_{mb}) I_m \end{cases}$$
(1)

where $I = I_a + I_b + I_m$. Note that $r_i(x) = \beta_i(x) - \alpha$ is the malthusian growth rate of the single-adapted genotype *i* (with $i \in \{a, b\}$) and $r_m = \beta_m - \alpha$ is the malthusian growth rate of the multi-adapted genotype *m*, when most of the hosts are uninfected (i.e. at the edge of the epidemic). Yet, when the pathogen population starts to increase locally the density of uninfected hosts drops and decreases the transmission opportunities as in classical epidemiological models with direct-transmission (see also (Débarre, Lenormand, and Gandon 2009; Griette, Raoul, and Gandon 2015). This drop in host density would be even stronger if host fecundity was not able to compensate host mortality (the total density of the host population would drop due to the spread of the pathogen). For simplicity, however, we restrict our analysis to the case where S(t, x) + I(t, x) remains constant and equal to K.

In the following we study the speed of spreading epidemics in a spatially heterogeneous environment as a function of (i) the period of the spatial fluctuation in the composition of the host population, (ii) the transmission rates of the different genotypes in the different habitats. We first consider the spread of single genotypes before analysing the effect of mutations among genotypes on the speed of a polymorphic pathogen population. Finally, we explore the effect of demographic stochasticity on the speed of monomorphic and polymorphic epidemics spreading in heterogeneous environments.

122 **3** Results

123 3.1 The speed of a monomorphic pathogen population

The multi-adapted genotype m does not "feel" the spatial heterogeneity of host population. When such a genotype is introduced in the host population and if we assume no mutation ($\mu_{ma} = \mu_{mb} = 0$) the above system reduces to the spread of a single pathogen in a uniform environment. The pathogen population spreads as a travelling wave with a speed equal to (Griette, Raoul, and Gandon 2015; Osnas, Hurtado, and Dobson 2015; Shigesada and Kawasaki 1997):

$$c_m = 2\sqrt{\sigma r_m}.\tag{2}$$

The analysis of the speed of a single-adapted genotype $i \in \{a, b\}$ is more challenging because the 129 growth rate of the pathogen varies periodically in space between $r_i(x) = r$ (when the genotype is 130 adapted to the host in x) and $r_i(x) = -r$ (when the genotype is not adapted to the host in x). It 131 is possible to derive good approximations for the speed of the epidemic in two limit cases (Hamel, 132 Fayard, and Roques 2010; Hamel, Nadin, and Roques 2011), namely when L is small and when L is 133 large. When the period of the fluctuation of the environment is very small (i.e. $L \rightarrow 0$) the grain 134 of the environment is so small that the growth rate of the pathogen is equal to the average growth 135 rate in the two habitats: $\overline{r} = \frac{r+(-r)}{2} = 0$. In contrast, when the period of the fluctuation is large 136 the pathogen will move very fast when it is adapted to the host and it will slow down when the host 137 resistance reduces its transmission rate. In the limit when $L \to \infty$ the speed reaches an asymptote 138

139 that can be described explicitly. We then get, for $i \in \{a, b\}$,

$$c_i \sim 0 \quad \text{when } 0 < L \ll 1, \quad c_i \sim \left(\frac{2}{\sqrt{3}}\right)^{3/2} \sqrt{r} \quad \text{when } 1 \ll L.$$
 (3)

Moreover the speed of the single-adapted genotype epidemic increases with L, the period of the spatial fluctuation of the environment (**Fig. 2**).

¹⁴² 3.2 The speed of a polymorphic pathogen population

Before considering the full system (with the 3 pathogen genotypes: a, b and m) we examine the 143 dynamics of a coalition of two single-adapted genotypes (a and b) each adapted to distinct types 144 of hosts. When the mutation rates are very low (i.e. $\mu_{aj} = \mu_{bj} \approx 0$) we recover the result of a 145 monomorphic population (red line in Fig. 2). However, numerical simulations with a fixed mutation 146 rate μ between single-adapted genotypes indicate that increasing the mutation rate has a complex effect 147 on the speed of the polymorphic population (Fig. 2). When L is small, increasing the mutation rate 148 has only a weak effect on epidemic speed because the environment changes so fast that both specialist 149 genotypes are almost equifrequent. For intermediate values of L, the size of the area populated by 150 a single host type allows the adapted genotype to outcompete the other genotype and to take up 151 some speed. Hence, the composition of the epidemic fluctuates between the two specialist genotypes 152 and a higher mutation rate speeds up the emergence of this locally adapted genotype and increases 153 the propagation speed. For larger values of L, however, this effect is dominated by the detrimental 154 emergence of ill-adapted mutants (*mutation load*) that slows down the propagation within an area 155 populated by a single host type. Hence, the composition of the pathogen population at the front of 156 the epidemic depends on the balance between local selection, mutation and L which measures the 157 amount of spatial heterogeneity. We show in the **Supplementary Information** (section 1.2.1) that 158 there is a threshold value L_c below which the whole epidemic can be driven by a single specialist: 159

$$L_c \sim \frac{2\sqrt{2}}{3^{3/4} - \sqrt{2}} \sqrt{\frac{\sigma}{r}} \ln\left(\frac{\sqrt{\sigma r}}{\mu}\right). \tag{4}$$

When $L < L_c$ the propagation of each specialist is independent because they can move through the "bad habitat" by diffusion. In contrast, when $L > L_c$ the bad habitat slows down the spread of the maladapted specialist and the coalition of two specialists is faster than a single specialist because they "pass the baton" when they move to a different habitat. The composition of the pathogen population at the front of the epidemic fluctuates between the two specialist genotypes. Higher mutation rates speed up the epidemic because mutation speeds up the switch between the two specialists at the tip of the front. Note, however, that high mutation rates generate a mutation load when $L \gg L_c$ via the recurrent introduction of a single-adapted genotype unable to infect the local host type. This is why the maximal speed of the coalition of single-adapted genotypes can never reach the speed of a universally adapted pathogen $(c_{a+b} < 2\sqrt{\sigma r} \text{ in Fig. 2}).$

When we assume a fixed mutation rate μ among the three pathogen genotypes, the epidemic 170 spreads faster than epidemics where only the coalition of two specialists is present, provided the 171 period of the fluctuation is small (Fig. 3). Indeed, when L is small the multi-adapted genotype m172 outpaces the single-adapted genotypes at the front of the epidemic (Fig. 3). In contrast, when L173 is large, the multi-adapted genotype is outcompeted by the coalition of the two specialists because 174 we assume the maximal growth rate r of the specialits is higher than the growth rate r_m of the 175 generalist (in particular when the mutation rate between single-adapted genotypes is large enough). 176 Increasing the mutation rate tends to lower the speed of the epidemic when L is small or very large. 177 because mutations reintroduce maladapted genotypes and build up the mutation load (Fig. 4). For 178 intermediate values of L, however, increasing the mutation rate can increase the speed of the pathogen 179 spread, by speeding up the propagation of a the coalition of specialists a and b (Fig. 4). This is due 180 to the beneficial effects of mutations on the speed of the coalition of two single-adapted genotypes 181 that we discussed above (**Fig. 2**). 182

183 3.3 The speed of stochastic epidemics

The above results rely on the assumption that the deterministic model we are using provides a good 184 description of the spread of a pathogen epidemics. Yet, the front of the epidemic is driven by a small 185 number of infections. The finite nature of the pathogen population at the edge of the epidemics 186 yields demographic stochasticity and is expected to slow down its spread (Brunet and Derrida 1997; 187 Griette, Raoul, and Gandon 2015; Mueller, Mytnik, and Quastel 2011; Snyder 2003). In the following 188 we explore the effect of stochasticity using an individual-based model that takes into account the 189 finite number N of hosts at each spatial location. The individual transitions between the different 190 states of the hosts are described by a list of random events (transmission, mutation, death; see the 191 Supplementary Information section 2.1 for a detailed description of the individual-based model). 192 As expected, this stochastic model converges to the above deterministic model when N is assumed 193 to be very large. To study the effect of demographic stochasticity on epidemic spread we performed 194 simulations with our individual-based model and measured the average speed on a long time interval 195 after the influence of the initial condition is lost. 196

¹⁹⁷ First, we discuss the speed of monomorphic epidemics in the absence of mutations. The speed of

the multi-adapted genotype is decreased by the effect of stochasticity but remains very close to the 198 deterministic approximation (see Brunet and Derrida 1997; Griette, Raoul, and Gandon 2015). The 199 magnitude of this drop is expected to be proportional to $\left(\ln\left(\frac{N}{\delta x}\right)\right)^{-2}$, where $\frac{N}{\delta x}$ represents the number 200 of hosts per unit of space. In contrast, the speed of the single-adapted specialist is dramatically 201 altered by stochasticity (Fig. 3). This speed is always lower than the speed of the deterministic 202 approximation but, when L is large the speed can drop abruptly to zero which indicates that the 203 pathogen cannot spread any more. Indeed, when the period of the fluctuation of the environment 204 reaches a threshold value $L_e \sim \frac{4}{3} \sqrt{\frac{\sigma}{r}} \ln\left(\frac{N}{\delta x}\right)$ the pathogen cannot cross the unfavourable habitat (see 205 Supplementary Information, section 2.2.2). In particular, the pathogen is very likely to go extinct 206 in the unfavourable habitat when the population size is small, the diffusion rate is limited and its 207 growth rate is very negative (remember that we assume the growth rate to be -r in the unfavourable 208 habitat). Note that this critical period L_e only increases logarithmically with the population size N, 209 so that this "blocking effect" can be observed even with relatively large population sizes. This explains 210 why the propagation speed of a single-adapted genotype is maximised for intermediate values of L. 211 In the deterministic approximation, in contrast, the pathogen can always cross unfavourable habitats 212 because extinctions do not occur and the speed of epidemic spread increases monotonically with L. 213 Second, if we allow some mutation between the two single-adapted genotypes, the epidemic can 214 cross those unfavorable environments because mutations will rescue pathogen populations when L > L215

 L_e . Consequently, increasing mutation rates can have a dramatic impact on the speed of epidemics when L is large (**Fig. 4**). Finally, when we allow the mutation between the three different genotypes, the speed of the epidemics is close to (but lower than) the deterministic approximation, and this speed can decrease when $L > L_e$ and the mutation rates are small enough (**Fig. 4**). As pointed out above, the magnitude of this effect on the reduction of the epidemic speed is of the order $(\ln(N))^{-2}$ when Nis large enough.

222 3.4 Pathogen diversity far behind the epidemic front

In the previous sections we focused on the speed and the composition of the pathogen population at the edge of the epidemic. Next, we characterise the composition of the pathogen population far behind the front, when it reaches an endemic equilibrium. Note that the composition of the pathogen population behind the front is much less sensitive to the effect of demographic stochasticity because at the endemic equilibrium, the number of pathogens present is much larger than at the front of the epidemics, diminishing greatly the risk of genotype extinctions. Hence, we do not need to distinguish the deterministic and stochastic models in this section. Three cases can be observed (**Fig. 5**): (i) The multi-adapted genotype dominates: If both the cost of being multi-adapted (i.e. $r - r_m$) and L are low, the generalist strategy outcompetes the specialists and goes to fixation.

(ii) The coalition of specialist genotypes dominates: When both the cost of being multi-adapted (i.e. $r - r_m$) and L are large, the coalition of specialists outcompetes the generalist strategy.

(iii) The three genotypes coexist: The coexistence between the three different genotypes is also possible for a range of parameter values when both r_m and L are relatively large. Indeed, as pointed by (Débarre and Lenormand 2011), a generalist strategy can outcompete specialists at the interface between habitats.

238 3.5 Five epidemic profiles

The above analysis shows how the composition of the pathogen population is dominated by different 239 genotypes at the edge and behind the front of the epidemic. Indeed, even if all genotypes are reintro-240 duced locally by mutation, the spatial variability of the environment and the spread of the population 241 affects the relative competitive abilities of the different genotypes at different locations. In particular, 242 when we vary both the period of host heterogeneity L and the growth rate r_m of the multi-adapted 243 genotype, we can distinguish five different profiles of epidemics (**Fig. 5**). Interestingly, we identify 244 an epidemic type (marked by III in Fig. 5, see also Fig. 6) where the multi-adapted genotype m245 drives the spread of the epidemic but is outcompeted later on by the coalition of the two specialists 246 (single-adapted genotypes a and b). In other words, the analysis of the transitory dynamics reveals 247 conditions where the multi-adapted genotype is able to emerge, taking advantage of the presence of 248 numerous uninfected host populations, even though specialized strategies are better competitors once 249 the epidemics has developed and many hosts have been infected. 250

We recover the same five epidemic profiles with finite host population sizes (**Fig. 5**) but demographic stochasticity affects the genetic diversity at the front of the epidemic where the size of the pathogen population is reduced. Single-adapted genotypes are most sensitive to the influence of stochasticity because these specialized genotypes can reach very low density in unfavourable habitats. The multi-adapted genotype m benefits from the influence of this demographic stochasticity (compare the size of epidemic type marked by **III** in the deterministic and stochastic cases illustrated by **Fig. 5**).

²⁵⁷ 4 Discussion

Our study provides a comprehensive analysis of the evolution of pathogen specialization in a spreading epidemics. Our model allows us to examine both the long-term evolutionary outcome far behind the front of the epidemic, and the transient evolution taking place at the front of the epidemic. We

recover the classical result of previous evolutionary analyses showing that the long-term evolutionary 261 outcome depends on the balance between spatial heterogeneity and the amount of migration among 262 habitats. Larger patches of homogeneous habitats favor the coalition of locally adapted specialists 263 in each habitat, but migration tends to favor generalist strategies able to cope with a diversity of 264 habitats (Christiansen 1975; Day 2000; Débarre and Gandon 2010; Débarre, Ronce, and Gandon 2013; 265 Mirrahimi and Gandon 2020). We also recover the possibility to maintain the coexistence of specialists 266 and generalist strategies when the generalist can be stably maintained at the interface between habitats 267 (Débarre and Lenormand 2011). Interestingly, our analysis of the transient evolutionary dynamics of 268 the pathogen in a spreading epidemic reveals that the composition of the pathogen population can 269 be very different at the front of the epidemic. Indeed, even if the local composition of the host 270 population does not change in time, the pathogen present at the front of the epidemic experiences 271 temporal fluctuations of the environment. Frequent temporal fluctuations favor the generalist strategy 272 because, in spite of its constitutive fitness cost (i.e. $r_m < r$ in our model), the generalist strategy does 273 not feel the heterogeneity of the environment. Consequently, we show that multi-adapted pathogens 274 are expected to drive the spread of epidemics in finely grained environments. In contrast, when the 275 spatial fluctuations are larger, the coalition of specialists is expected to drive the epidemics. Indeed, 276 even if the transition between the two habitats can slow down the average speed of a coalition of 277 specialists, the speed of each specialist is maximized when they are locally adapted. Contrasting the 278 composition of the pathogen population at the edge and at the back of the epidemic allowed us to 279 identify five different types of epidemic profiles in Fig. 5. This figure shows that the coexistence of 280 specialists and generalists strategies is promoted by a lower fitness of the multi-adapted genotype and 281 a larger period of host heterogeneity. In general we find that the speed of the epidemic is increased 282 with larger period of host heterogeneity but, as discussed below, these results are modulated by the 283 pathogen mutation rates and by the amount of demographic stochasticity. 284

We found that mutation among pathogen genotypes is a double edged sword: (i) it allows the 285 pathogen to acquire adaptive mutations but (ii) it can also produce a mutation load with the recur-286 rent introduction of locally maladapted genotypes. The balance between these two effects depends 287 on the heterogeneity of the environment which, in turn, depends on the ratio between the period L288 of the fluctuation of the environment and the diffusion coefficient σ . The beneficial effect of a higher 289 mutation rate is maximal for intermediate levels of this ratio. Indeed, it is not profitable for the 290 pathogen population to mutate often when the environment keeps changing (i.e., $L \sim 0$) or when the 291 environment changes very slowly (i.e., $L \to \infty$). Several earlier studies obtained similar conclusions 292 in non-spatial models where it is possible to show that there is an optimal stochastic switching rate 293

²⁹⁴ between specialized phenotypes that maximizes the growth rate of a population in a fluctuating en-²⁹⁵ vironment (Kussell and Leibler 2005; Lachmann and Jablonka 1996). In all these different scenarios, ²⁹⁶ the introduction of genetic variation provides a way to "pass the baton" between different specialist ²⁹⁷ genotypes and allows the population to exploit more efficiently a fluctuating environment.

As expected from earlier theoretical studies (Brunet and Derrida 1997; Griette, Raoul, and Gandon 298 2015; Mueller, Mytnik, and Quastel 2011; Snyder 2003), demographic stochasticity lowers the speed 299 of the epidemic spread. Most of the results of the deterministic model hold in finite host populations. 300 The only notable exception occurs when large values of L can prevent the spread of single-resistance 301 genotypes. The input of new mutations may then provide a way to adapt to the new host type. Hence 302 the speed of pathogen epidemics may be constrained by both the stochastic nature of the demographic 303 process and the stochastic nature of the mutation events occurring at the edge of the epidemic. Several 304 earlier studies have shown how the increased intensity of genetic drift in expanding populations could 305 result in an "expansion load" due to the accumulation of deleterious mutations (Hallatschek and 306 Nelson 2010; Peischl, Kirkpatrick, and Excoffier 2015). In our model, however, deleterious mutations 307 at some location (e.g. genotype a in host type B) are adaptive at other locations (e.g. in host type A). 308 It would be interesting to study the effects of finite population size in a more realistic model allowing 309 for the accumulation of unconditionally deleterious mutations. 310

Our models can be used to make practical recommendations regarding the manipulation of the 311 spatial structure of the host population to limit the speed of pathogen epidemics. The spatial structure 312 of the host population can be manipulated by mixing hosts with different levels of resistance to 313 the pathogen. This variation in host resistance can either be due to genetic heterogeneity among 314 (e.g. resistant crop varieties), to immunological heterogeneity (e.g. vaccination) or other therapeutic 315 interventions (e.g. the use of drugs against the pathogen). Earlier studies have analysed the impact 316 of the local manipulation of the heterogeneity of the environment on the adaptation of pests and 317 pathogens (Comins 1977; Débarre, Bonhoeffer, and Regoes 2007; Lenormand and Raymond 1998; 318 Park et al. 2015; Raymond 2019). In particular these models have determined the critical area size of 319 host resistance above which adaptation to the host does not occur because local selection is swamped 320 by the influence of migration. The present study expands these earlier studies that focused on the 321 migration-selection equilibrium and examines transient dynamics of adaptation in the presence of 322 two types of host resistance. Hence, our analysis may be particularly relevant in agriculture where 323 multiple resistance varieties may be used to limit pathogen spread (Djidjou-Demasse, Moury, and 324 Fabre 2017; Rimbaud et al. 2018a,b, 2021). If the objective is to limit the speed of the epidemic 325 spread, a lower value of L should be recommended. Lower L values imply that a spreading epidemics 326

is exposed to a more variable environment. This prevents the pathogen to specialize to a specific 327 environment and, consequently, to speed up in a favourable environment. Interestingly, fine-scale 328 environmental heterogeneity (low L values) are also expected to reduce the probability of pathogen 329 emergence (Chabas et al. 2018). This fine-scale heterogeneity, however, may promote the spread of 330 generalist and multi-adapted pathogens. Those generalist pathogens are likely to spread more slowly 331 because of the potential fitness cost associated with the acquisition of additional mutations. But 332 additional compensatory mutations (not considered in our model) may restore the competitivity of 333 generalist pathogens against specialist pathogens. In other words, the optimal deployment of control 334 measures in space varies with the forecast horizon. Our model helps clarify the consequences of these 335 interventions on the short term epidemiological dynamics (the speed of the spreading epidemic) as 336 well as the evolutionary dynamics of the pathogen population. 337

Several experimental studies have monitored and quantified the spread and the evolution of a 338 bacteria in laboratory conditions (Baym et al. 2016; Deforet et al. 2019). In particular, the MEGA-339 plate experiment of Baym et al followed the spread of *Escherichia coli* in a spatially heterogeneous 340 environment characterised by increasing concentrations of antibiotics. This fascinating experiment 341 allowed to visualize pathogen spread and evolution in real time. This experimental procedure could 342 be used to test some of our predictions. For instance, we could monitor the influence of the scale of 343 spatial heterogeneity with a manipulation of the parameter L in the MEGA-plate. We hope that the 344 present theoretical framework may stimulate an experimental validation of our theoretical predictions 345 using experimental evolution of microbes in spatially heterogeneous environments. 346

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352 Author contributions

QG carried out the simulations and made the figures, MA contributed to the analysis of the model, GR derived the approximations of the threshold values of habitat size, SG wrote the first draft of the manuscript, and all authors contributed to the revisions of the manuscript. The project was initiated by GR and SG, and all authors contributed to the development of the final version of the model.

357 Data accessibility

³⁵⁸ This is a theoretical study and no data were used.

359 References

- Baym, M. et al. (2016). Spatiotemporal microbial evolution on antibiotic landscapes. Science 353.6304,
- ³⁶¹ pp. 1147–1151.
- Berestycki, H., F. Hamel, and L. Roques (2005a). Analysis of the periodically fragmented environment
 model. I. Species persistence. J. Math. Biol. 51.1, pp. 75–113.
- (2005b). Analysis of the periodically fragmented environment model. II. Biological invasions and pulsating travelling fronts. J. Math. Pures Appl. (9) 84.8, pp. 1101–1146.
- Brunet, E. and B. Derrida (1997). Shift in the velocity of a front due to a cutoff. *Physical Review E*56.3, p. 2597.
- Chabas, H. et al. (2018). Evolutionary emergence of infectious diseases in heterogeneous host popula tions. *PLoS biology* 16.9, e2006738.
- Christiansen, F. B. (1975). Hard and soft selection in a subdivided population. The American Naturalist 109.965, pp. 11–16.
- Comins, H. N. (1977). The management of pesticide resistance. Journal of Theoretical Biology 65.3,
 pp. 399–420.
- Day, T. (2000). Competition and the effect of spatial resource heterogeneity on evolutionary diversi fication. *The American Naturalist* 155.6, pp. 790–803.
- Débarre, F. and S. Gandon (2010). Evolution of specialization in a spatially continuous environment.
 Journal of evolutionary biology 23.5, pp. 1090–1099.
- ³⁷⁸ Débarre, F. and T. Lenormand (2011). Distance-limited dispersal promotes coexistence at habitat ³⁷⁹ boundaries: reconsidering the competitive exclusion principle. *Ecology Letters* **14.3**, pp. 260–266.
- Débarre, F., T. Lenormand, and S. Gandon (2009). Evolutionary epidemiology of drug-resistance in
 space. *PLoS Computational Biology* 5.4, e1000337.
- ³⁸² Débarre, F., O. Ronce, and S. Gandon (2013). Quantifying the effects of migration and mutation
- on adaptation and demography in spatially heterogeneous environments. Journal of Evolutionary
 Biology 26.6, pp. 1185–1202.
- Deforet, M., C. Carmona-Fontaine, K. S. Korolev, and J. B. Xavier (2019). Evolution at the edge of expanding populations. *The American Naturalist* **194.3**, pp. 291–305.

- ³⁸⁷ Djidjou-Demasse, R., B. Moury, and F. Fabre (2017). Mosaics often outperform pyramids: insights ³⁸⁸ from a model comparing strategies for the deployment of plant resistance genes against viruses in ³⁸⁹ agricultural landscapes. *New Phytologist* **216.1**, pp. 239–253.
- Débarre, F., S. Bonhoeffer, and R. R. Regoes (2007). The effect of population structure on the emergence of drug resistance during influenza pandemics. *Journal of The Royal Society Interface* 4.16,
 pp. 893–906.
- Fisher, R. A. (1937). The wave of advance of advantageous genes. Annals of eugenics 7.4, pp. 355–369.
- Gilligan, C. A. (2008). Sustainable agriculture and plant diseases: an epidemiological perspective.
 Philosophical Transactions of the Royal Society B: Biological Sciences 363.1492, pp. 741–759.
- Griette, Q., G. Raoul, and S. Gandon (2015). Virulence evolution at the front line of spreading
 epidemics. *Evolution* 69.11, pp. 2810–2819.
- Hallatschek, O. and D. R. Nelson (2010). Life at the front of an expanding population. Evolution 64.1,
 pp. 193–206.
- Hamel, F., J. Fayard, and L. Roques (2010). Spreading speeds in slowly oscillating environments. *Bull. Math. Biol.* **72.5**, pp. 1166–1191.
- Hamel, F., G. Nadin, and L. Roques (2011). A viscosity solution method for the spreading speed
 formula in slowly varying media. *Indiana Univ. Math. J.* 60.4, pp. 1229–1247.
- Kolmogorov, A. N., I. G. Petrovsky, and N. S. Piskunov (1937). Étude de l'équation de la diffusion
 avec croissance de la quantité de matière et son application à un problème biologique. *Bull. Univ.*
- 406 Etat Moscou Sér. Inter. A 1, pp. 1–26.
- Kot, M., M. Lewis, and P. van den Driessche (1996). Dispersal Data and the Spread of Invading
 Organisms. *Ecology* 77.7, pp. 2027–2042.
- Kussell, E. and S. Leibler (2005). Phenotypic diversity, population growth, and information in fluctuating environments. *Science* **309.5743**, pp. 2075–2078.
- Lachmann, M. and E. Jablonka (1996). The inheritance of phenotypes: an adaptation to fluctuating
 environments. *Journal of theoretical biology* 181.1, pp. 1–9.
- Lenormand, T. and M. Raymond (1998). Resistance management: the stable zone strategy. *Proceedings*of the Royal Society of London. Series B: Biological Sciences 265.1409, pp. 1985–1990.
- Levins, R. (1968). Evolution in changing environments: some theoretical explorations. 2. Princeton
 University Press.
- ⁴¹⁷ McLeod, D. V., L. M. Wahl, and N. Mideo (2021). Mosaic vaccination: how distributing different
 ⁴¹⁸ vaccines across a population could improve epidemic control. *Evolution Letters* 5.5, pp. 458–471.

- ⁴¹⁹ Mikaberidze, A., B. A. McDonald, and S. Bonhoeffer (2015). Developing smarter host mixtures to ⁴²⁰ control plant disease. *Plant Pathology* **64.4**, pp. 996–1004.
- Mirrahimi, S. and S. Gandon (2020). Evolution of specialization in heterogeneous environments: equilibrium between selection, mutation and migration. *Genetics* 214.2, pp. 479–491.
- Mueller, C., L. Mytnik, and J. Quastel (2011). Effect of noise on front propagation in reaction-diffusion
 equations of KPP type. *Invent. Math.* 184.2, pp. 405–453.
- Mundt, C. C. (2002). Use of multiline cultivars and cultivar mixtures for disease management. Annual
 review of phytopathology 40.1, pp. 381–410.
- ⁴²⁷ Osnas, E. E., P. J. Hurtado, and A. P. Dobson (2015). Evolution of pathogen virulence across space
 ⁴²⁸ during an epidemic. *The American Naturalist* 185.3, pp. 332–342.
- Park, A. W., J. Haven, R. Kaplan, and S. Gandon (2015). Refugia and the evolutionary epidemiology
 of drug resistance. *Biology Letters* 11.11, p. 20150783.
- Parvinen, K. and M. Egas (2004). Dispersal and the evolution of specialisation in a two-habitat type
 metapopulation. *Theoretical Population Biology* 66.3, pp. 233–248.
- Peischl, S., M. Kirkpatrick, and L. Excoffier (2015). Expansion load and the evolutionary dynamics of
 a species range. *The American Naturalist* 185.4, E81–E93.
- Perkins, T., B. Phillips, M. Baskett, and A. Hastings (2013). Evolution of dispersal and life history
 interact to drive accelerating spread of an invasive species. *Ecol Lett* 16, pp. 1079–1087.
- Raymond, B. (2019). Five rules for resistance management in the antibiotic apocalypse, a road map
 for integrated microbial management. *Evolutionary applications* 12.6, pp. 1079–1091.
- 439 Rimbaud, L., J. Papaix, L. G. Barrett, J. J. Burdon, and P. H. Thrall (2018a). Mosaics, mixtures, ro-
- tations or pyramiding: What is the optimal strategy to deploy major gene resistance? *Evolutionary*
- ⁴⁴¹ Applications **11.10**, pp. 1791–1810.
- Rimbaud, L., J. Papaix, J.-F. Rey, L. G. Barrett, and P. H. Thrall (2018b). Assessing the durability and efficiency of landscape-based strategies to deploy plant resistance to pathogens. *PLoS computational biology* 14.4, e1006067.
- Rimbaud, L. et al. (2021). Models of plant resistance deployment. Annual Review of Phytopathology
 59, pp. 125–152.
- Shigesada, N. and K. Kawasaki (1997). Biological invasions: theory and practice. Oxford University
 Press, UK.
- ⁴⁴⁹ Skellam, J. (1951). Random dispersal in theoretical populations. *Biometrika* **38**, pp. 196–268.
- Snyder, R. E. (2003). How demographic stochasticity can slow biological invasions. *Ecology* 84.5,
 pp. 1333–1339.

- ⁴⁵² Wei, W. and S. M. Krone (2005). Spatial invasion by a mutant pathogen. *Journal of theoretical biology*
- **236**.3, pp. 335–348.
- Wilson, D. S. and J. Yoshimura (1994). On the coexistence of specialists and generalists. *The American Naturalist* 144.4, pp. 692–707.

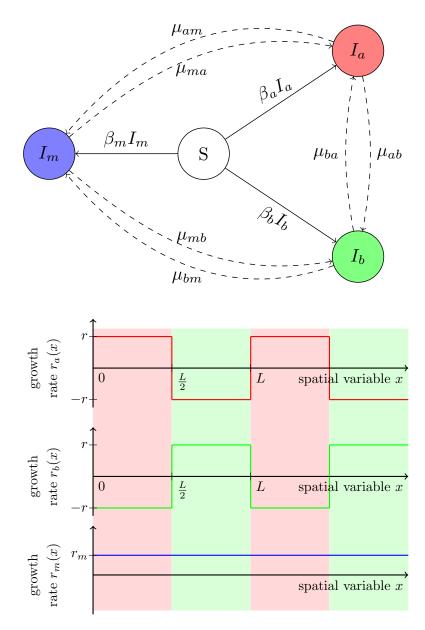


Figure 1: Schematic presentation of the evolutionary epidemiology model and the spatial heterogeneity of the environment. Top figure: diagram of the compartimental model. S represents susceptible hosts, I_a (resp. I_b , I_m) represents hosts infected by single-adapted genotype a, which is only able to infect host type A (resp. single-adapted genotype b only able to infect host type B, and the multi-adapted pathogen able to infect both types of hosts). In dashed we have represented mutations that typically happen at a much lower rate than transmissions. Bottom figure: Values of the intrinsic growth rates $x \mapsto r_a(x) = \beta_a(x) - \alpha$, $x \mapsto r_b(x) = \beta_b(x) - \alpha$, $x \mapsto r_m = \beta_m - \alpha$ as a function of the spatial variable $x \in \mathbb{R}$, where, for $x \in (0, L)$, $\beta_a(x) = 2r \mathbb{1}_{(0, \frac{L}{2})}(x)$ while $\beta_b(x) = 2r \mathbb{1}_{(\frac{L}{2}, L)}(x)$, and $\alpha = r$. The maximal growth rate of the single-adapted genotypes is assumed to be higher than the growth rate of the multi-adapted genotype: $r \geq r_m$. The red (resp. green) area represents the locations $x \in \mathbb{R}$ where hosts of type A (resp. B) are present.

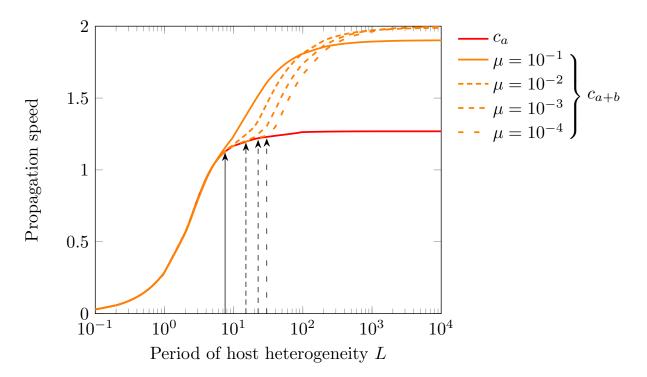


Figure 2: Impact of the mutation rate μ on the propagation speed of a coalition of the two specialist pathogen types, for the determinist model. We plot the speed c_a of a single specialist genotype (red line) and the speed c_{a+b} of a coalition of both specialist genotypes propagating together (orange lines) when $\mu_{ab} = \mu_{ba} = \mu$ (with $\mu_{am} = \mu_{bm} = 0$). The final values for c_a are extrapolated (from L = 2000 inclusive). The black arrows indicate the values of L_c for the different rates of mutation (see equation (4)). Parameters: $\sigma = 1$, r = 1, and the functions $\beta_a(x)$, $\beta_b(x)$, $r_a(x)$ and $r_b(x)$ are as in Fig. 1.

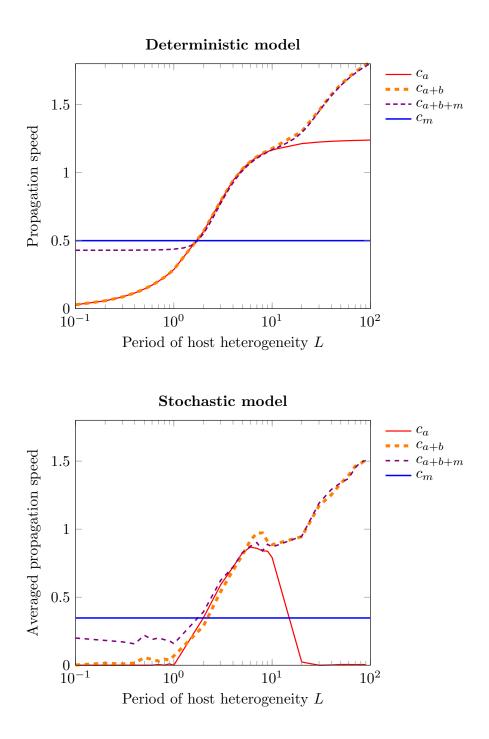


Figure 3: Propagation speed when only one specialist genotype is present (c_a) , when both specialist genotypes are present $(c_{a+b} \text{ with } \mu_{ab} = \mu_{ba} = \mu)$ and when all the three pathogen genotypes are present $(c_{a+b+m} \text{ with } \mu_{ij} = \mu, \forall i, j \in \{a, b, m\})$. Top figure: speed of the epidemic in the *deterministic model* (1) against the period *L* for the coalition of specialist genotypes (orange line: c_{a+b} with $\mu_{ab} = \mu_{ba} = \mu$), the multi-adapted genotype alone (blue line: c_m) and the full model with both the specialist genotypes and the multi-adapted genotype (purple line: c_{a+b+m} with $\mu_{ij} = \mu, \forall i, j \in \{a, b, m\}$). Bottom figure: speed of the epidemic in the *stochastic model* with N = 100and $\delta x = 0.1$. Parameters: r = 1, $r_m = \frac{1}{16}$, $\sigma = 1$, $\mu = 0.01$, $\beta_m = 1 + \frac{1}{16}$, and the functions $\beta_a(x)$, $\beta_b(x)$, $r_a(x)$ and $r_b(x)$ are as in Fig. 1.

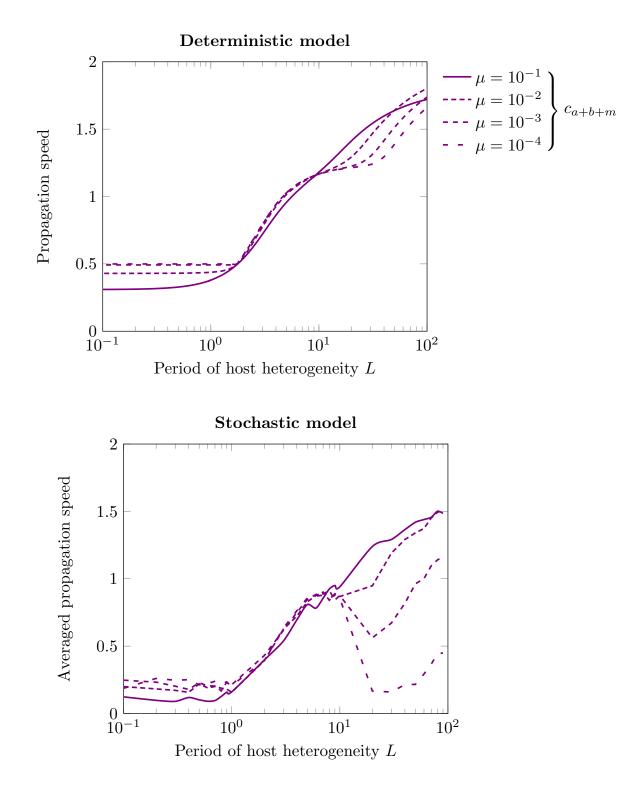


Figure 4: Effect of the mutation rate μ on the propagation speed of the epidemics when all three pathogen types are present $(c_{a+b+m} \text{ with } \mu_{ij} = \mu, \forall i, j \in \{a, b, m\})$. Top figure: *deterministic model*. Bottom figure: *stochastic model* with N = 100 and $\delta x = 0.1$. Parameters: $\sigma = 1$, $r_m = \frac{1}{16}$, r = 1, $\beta_m = 1 + \frac{1}{16}$, and the functions $\beta_a(x)$, $\beta_b(x)$, $r_a(x)$ and $r_b(x)$ are as in Fig. 1.

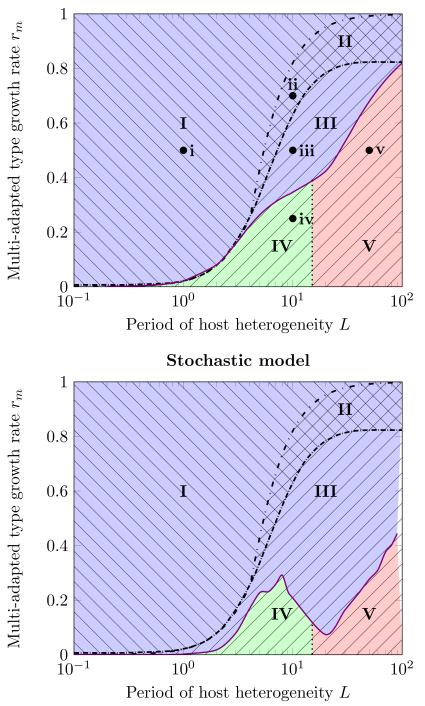
Multi-adapted type at the edge

Multi-adapted type behind the front

WWW Multi-adapted and specialist types behind the front

 \square A single specialist type at the edge \blacksquare Both specialist types behind the front

Both specialist types at the edge



Deterministic model

Figure 5: The five epidemic profiles. Composition of the population at the edge of the front (colors), and behind the front (hatches), as a function of r_m and L with $\mu_{ij} = \mu, \forall i, j \in \{a, b, m\}$. See also Fig. 6 for the description of these different epidemic profiles obtained with the parameters noted **i** to **v** in the top figure . Top figure: *deterministic model*. Bottom figure: *stochastic model* with N = 100 and $\delta x = 0.1$. Parameters: $\sigma = 1, \mu = 0.01, r = 1, \beta_m = 1 + r_m$, and the functions $\beta_a(x)$, $\beta_b(x), r_a(x)$ and $r_b(x)$ are as in Fig. 1.

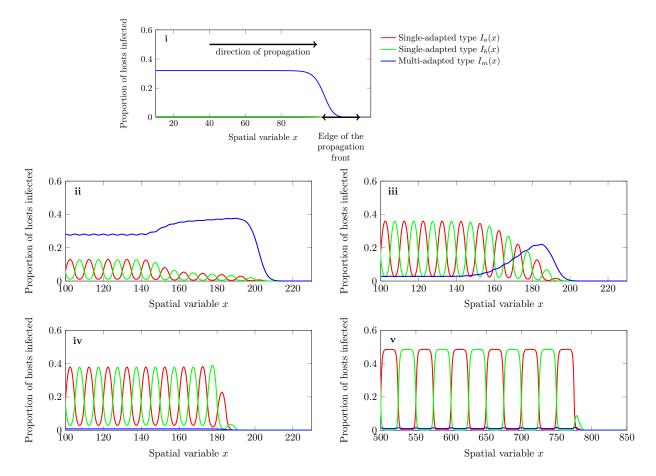


Figure 6: Composition of the pathogen population. For the parameters (L, r_m) noted i to v in Fig. 5: (1, 0.5), (10, 0.7), (10, 0.5), (10, 0.25), (50, 0.5). Other parameters r = 1, $\mu = 0.001$, $\beta_m = 0.5$, and the functions $\beta_a(x)$, $\beta_b(x)$, $r_a(x)$ and $r_b(x)$ are as in Fig. 1.